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БЕНТИЧЕСКИЕ ФОРАМИНИФЕРЫ КАК ИНДИКАЦИОННОЙ ИНСТРУМЕНТ БИОСТРАТИГРАФИИ И ПАЛЕОЭКОЛОГИИ ПАЧКИ ГУРИ (ФОРМАЦИЯ МИШАН), БАНДАР-АББАС, ЮЖНЫЙ ИРАН

BENTHIC FORAMINIFERA AS A TOOL FOR INDICATION OF BIOSTRATIGRAPHY, AND PALEOECOLOGY OF THE GURI MEMBER (MISHAN FORMATION), BANDAR ABBAS, SOUTH IRAN

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Аннотация. Это исследование сосредоточено на биостратиграфии и палеоэкологии фораминифер пачки Гури (формация Мишан) к северо-западу от Бендер-Аббаса, к югу от Ирана и в бассейне Загрос (восточная провинция Тетиса). Пачка Гури в основном состоит из средне- и толстослоистых известняков, среднеслоистых мергелистых известняков и зеленого мергеля с прослоями средних коричневых известняков. В этом исследовании было установлено пять биозон бентосных фораминифер, включающих 21 вид бентосных фораминифер из 17 родов. Сравнение выявленных бентосных фораминиферовых биозон пачки Гури с таковыми в других частях мира обнаруживает близкое сходство с областью Тетис. Фораминиферы подтверждают ранний-средний миоцен пачки Гури. Семь морских высокоизвестковых видов остракод встречались только в мергелевых отложениях, но не в известняках. Биотические ассоциации, выявленные в этом исследовании, позволяют предположить, что карбонатная седиментация мишанской свиты от нижней части к верхней части процветала в условиях от субтропических до тропических, отражала мезотрофные и олиготрофные палео условия, где отлагала мягкие до твердых отложений на глубине воды от 20 до 80 м и откладывались в условиях нормальной солености воды, которая колеблется от 34 до 50 ‰.

Ключевые слова: биостратиграфия; палеоэкология; миоцен; Бассейн Загрос; мишанская свита; группа Гури; Фораминифера, Остракода, Бандар Аббас.

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Annotation. This study is focused on the foraminiferal biostratigraphy and paleoecology of the Guri Member (Mishan Formation) northwest of Bandar Abbas, south of Iran, and in the Zagros Basin (eastern Tethyan province). The Guri Member mainly consists of medium- to thick-bedded limestone, medium-bedded marly limestone, and green marl with intercalated medium, brown limestones. In this study, five benthic foraminiferal biozones were established, including 21 benthic foraminiferal species from 17 genera. A comparison of the identified benthic foraminiferal biozones of the Guri Member with those in other parts of the world reveals close similarity to the Tethyan realm. Foraminifera confirms the Early to middle Miocene for the Guri Member. Seven marine high calcareous rare Ostracoda species were found in marl sediments only, not in Limestone layers. The biotic associations identified in this study suggest that the carbonate sedimentation of the Mishan Formation from the lower part to the upper part thrived in subtropical to tropical environments, reflected mesotrophic to oligotrophic conditions, deposited soft to hard sediments at water depths that ranged from 20–80 m, and deposited in a normal water salinity environment that ranges from 34 to 50 ‰.

Keywords: biostratigraphy; Geology; Paleocology; Miocene; Zagros Basin; Foraminifera, Ostracoda, Mishan Formation; Guri Member; Bandar Abbas, Iran.

1 INTRODUCTION

The Mishan Formation has previously been called the Middle Fars, its usage is also extended into Fars province, thus replacing the names argillaceous group and Anguru Marl. The type of section was measured at the southwestern Gachsaran Oil Field at approximately 710 m thickness of marl and limestone. (James and Wynd, 1956), the sharp basal contact with gypsum of the Gachsaran Formation is accompanied

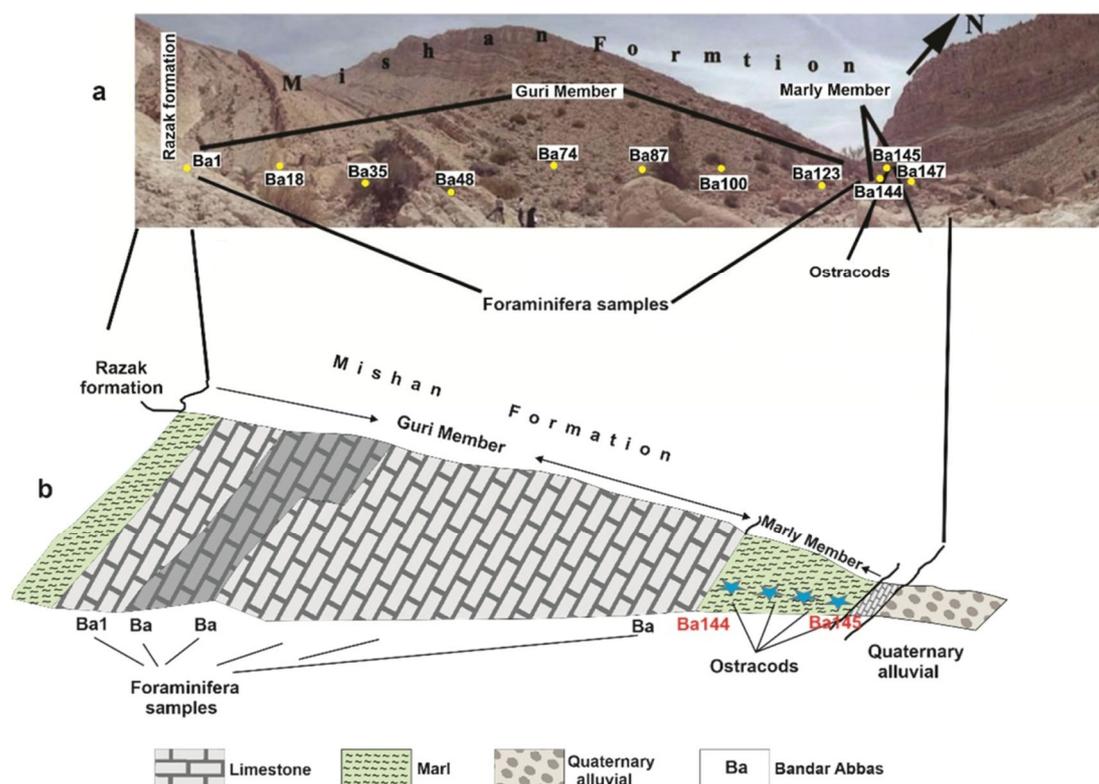


Fig. 3. Baghestan section, northwest of the Bandar Abbas Hinterland, south Iran is sampled the letter Ba1-Ba147 are the locations of the samples: a – field sampling; b – cross-section

Motiei, 1993; Goof et al., 1994; Homaiun-Zadeh, 2002; Alsouki et al., 2008). The Mishan Formation is equivalent to the Jerebi and Fatha formations in Iraq (Ghafor, 2004, 2010, 2011, 2014, 2015, 2022a; Ghafor and Ahmad, 2019, 2021; Ghafor and Muhammad, 2005, 2007, 2011; Ghafor and Najaflo, 2022; Ghafor et al. 2014; Ghafor et al. 2023; Muhammad and Ghafor, 2008). The main goal of the present study is to examine the biostratigraphy and paleoecology of the Mishan Formation in the studied section and correlate bio-events in the Tethyan realms.

2 GEOLOGICAL SETTING

According to Bahroudi, A., and Koyi Hemin, (2004), the Zagros Basin is defined by a 7–14 km thick succession of deposits over a region along the north–northeastern edge of the Arabian plate. This basin was part of the stable Gondwana supercontinent in the Paleozoic Era, a passive margin in the Mesozoic, and became a site of convergent orogeny in the Cenozoic. Sadeghi et al., (2009) clarified that the Zagros Fold-and-Thrust Belt of Iran is the result of Alpine orogenic events, which occurred in the Alp–Himalayan Mountain range. It extends in an NW–SE direction from eastern Turkey to the Strait of Hormoz in southern Iran. The tectonic activity of this area was entirely due to the convergence of the Arabian and Eurasian continents. Amirshahkarami et al. (2006), clarified that the Arabian and Iranian plates were part of the Eurasian plate and collided during the Mesozoic and Cenozoic periods, creating the Zagros folding belt. The Zagros Basin is a part of the middle section of the Tethyan realm and covers an area of nearly 500 km, with an NW-SE length of about 2300 km and a NE-SW width of 100–300 km. It mainly includes SW Iran, NE Iraq, NE Syria, and SE Turkey. This basin extends from SE Turkey to SW Iran and contains the provinces of Lorestan, Khuzestan, and Fars of the Zagros Basin (Motiei, 1995; Aghanabati, 2006). The Mishan Formation is exposed and present in most parts of the Zagros basin (Motiei 1993), but it's well developed in Hormozgan Province and the Bandar Abbas Hinterland. Bandar-Abbas Hinterland is in the southeastern part of the Zagros Mountains, and it has a different orientation than other parts of the sedimentary basin (Haynes and McQuilan, 1974). This hinterland is bounded by the Minab Fault on the east, the Zagros fold belt, which crosses the Persian Gulf on the south, and the Razak Fault on the north (Barzgar, 1981). Most of the structures are anticlines exposing Fars Group successions, which are thicker than the Fars Zone in the study area, and the eastern border is limited to the Minab fault in the southern border of the rest of Zagros simple fold zone (Aghanabati, 2004). Stöcklin, 1968, subdivided the Iranian Plateau into eight continental parts. (i.e., Zagros, Sanandaj Syrjan, Urmiah-Dokhtar, Central Iran, Alborz, Kopeh-Dagh, Lut, and Makran), based on the sedimentary sequence, structural setting, intensity of deformation, magmatism, and metamorphism, but (Aghanabati, 2004) subdivide the Zagros Zone into two subzones (i.e., Thrust Subzone and Folded Zagros)



based on geology characteristics. Folded Zagros Subzone has been subdivided into seven parts, including Lurestan, Khuzestan, Abadan Plain, Fars (external and interior), Bandar Abbas Hinterland, and a complex structure with metamorphic rock (Fig. 4). based on lateral facies variations, the Iranian Zagros fold-thrust belt is divided into different tectonostratigraphic domains that, from SE to NW, are the Fars Province (Eastern Zagros), the Khuzestan Province (Central Zagros), and the Lurestan Province (Western Zagros), respectively (Motiei, 1993). Hormozgan Province is in southern Iran and is part of the Zagros Folded Belt. This region is accompanied by NW-SE, W-E, and N-S trending simple anticlines and synclines with the very great thickness of Fars Group deposits (Gachsaran, Mishan, Aghajari, and Bakhtiari formations) and the presence of 118 salt plugs. So, for these specific features, Motiei, (1993) called this area the «Bandar Abbas Hinterland.»

The Zagros Fold-and-Thrust Belt of Iran is a result of the Alpine orogenic events in the Alpine–Himalayan Mountain range (Sadeghi et al., 2009). This basin was part of the stable Gondwana supercontinent in the Paleozoic Era, a passive margin in the Mesozoic Era, and a site of convergent orogeny in the Cenozoic Era (Bahroudi and Koyi, 2004). It extends in an NW–SE direction from eastern Turkey to the Strait of Hormoz in southern Iran. The tectonic activity of this area was entirely due to the convergence of the Arabian and Eurasian continents. (Aghanabati, 2004).

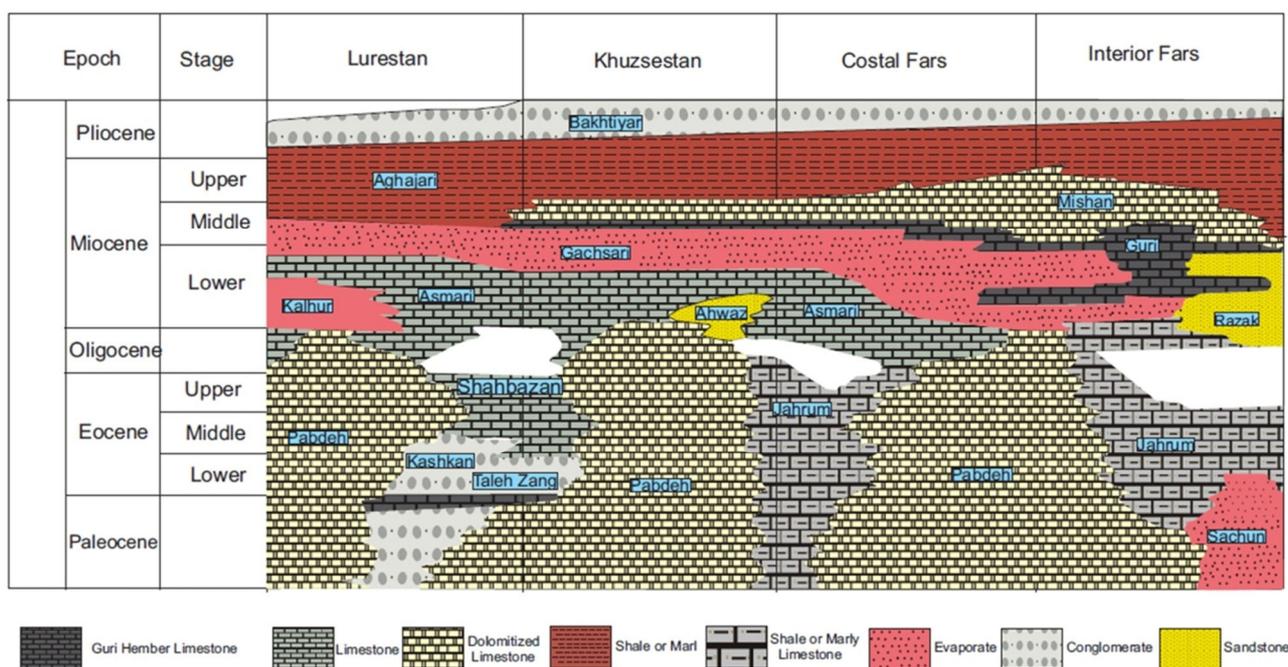


Fig. 4. Lithostratigraphic units of the Cenozoic of Zagros Fold-Thrust (modified from Gulf Petrolink, 1998)

3 MATERIALS AND METHODS

147 samples were collected at 0.5–3 m intervals from the Baghestan section of the Mishan Formation (496 m thick). 200 thin sections were prepared in the laboratory, and the thin sections were examined by using a polarized microscope. Foraminiferal identification and biostratigraphy were done by the foraminiferal classifications (Bakhtiar and Taheri 2010; Sirel et al., 2013; Serra-Kiel et al., 2016; Boudagher-Fadel 2018; Joudaki and Baghbani 2018; Moghaddam et al., 2019), and the studies of (Burchette and Wright, 1992; BouDagher-Fade,l 2008) are applied for palaeoecological interpretation. Seven marine high calcareous Ostracoda species found in marl sediments of Bandar-Abbas were prepared for SEM photography.

4 RESULTS AND DISCUSSION

4.1 Biostratigraphy

The biostratigraphy framework of the Mishan Formation was reported by Wynd, (1965) and developed by Adams et al., (1967). The studied section is characterized by shallow-benthic zones (SBZ) of the Early-Middle Miocene foraminifera related to these families: *Sortidae*, *Peneroplidae*, *Miliolidae*, *Rotalidae*, *Amphestedae*, *Hauerinidae*, *Alveolinidae*, *Meandrospiniidae*, *Elephinidae*, *Miogypsinidae*, *Miogypsinidae*, and A total of 21 species from 17 genera of LBF and 7 species of non- Foraminifera were identified, including algae (*Subterraniophyllum thomasi*, *Lithophyllum* sp., *Lithothamnium* sp.), bryozoans (*Tubucellaria* sp., *Onyococella* sp., *Memberanipora* sp.), molluscan macrofossils (bivalves and gastro From base to top, five biozones were recognized (Plates 1–3, Fig. 5 and 6). The presence of limited marine Miocene Ostracoda species confirms the biostratigraphy of foraminifera fauna collected from field outcrop samples. (Plate 4)

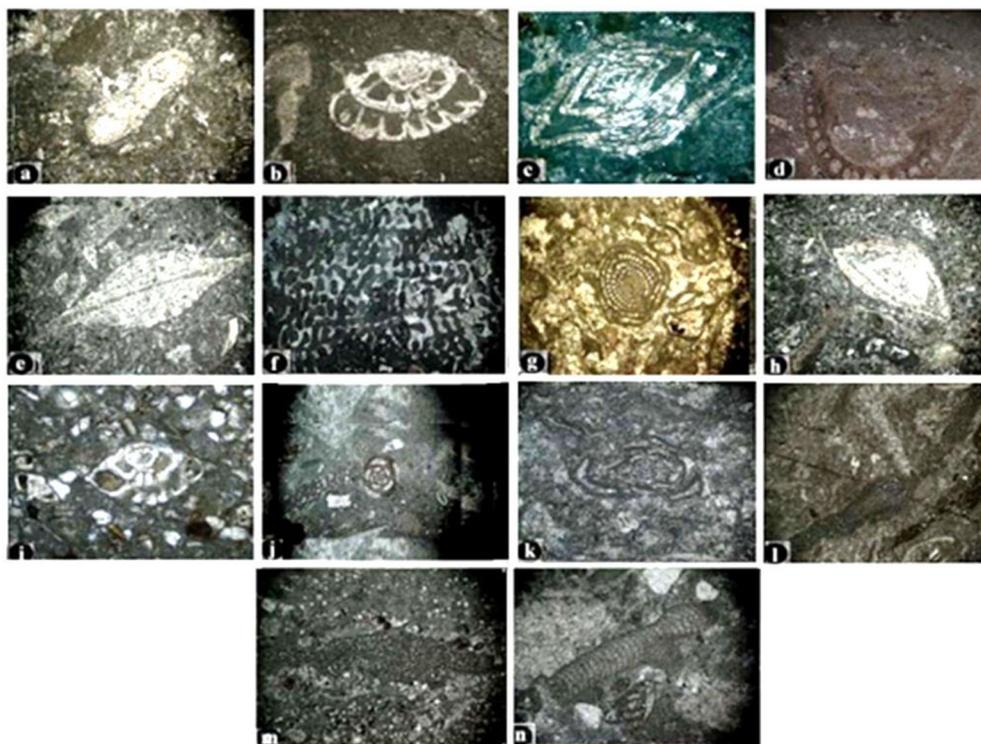


Plate-1. a. *Discorbis* sp. (10 X) sample no. 8; b. *Ammonia beccari* (Line, 1758), (10 X) sample no. 33; c. *Amphistegina lessoni* d'Orbigny in Guérin-Méneville, 1832, (10 X) sample no. 12.; d. *Memberanipora* sp. (4X) Sample no: 80; e. *Discocyclus* sp. (4X) sample no. 7; f. coral,), sample no. p.; g. *Borelis haueri* Azmi, 2010 (4X) sample no: 85; h. *Amphistegina radiat* (Fichtel & Moll, 1798) (10X) sample no. 132; i. *Ammonia stachi* d' Orbigny, 1826 (10X) sample no. 2; *Quinquiloculina* sp. (4X) sample no. 101. k. *Schlumbergerina* sp. (10 X) sample no: 10; l. *Archaias hensoni* Smout and Eames, 1958 (4X) Sample no. 36; m. *Archias kirkukensis* Henson, 1950 (4X) sample no. 17; n. *Subterraniophyllum thomasi* Elliott (4X) Sample no.10

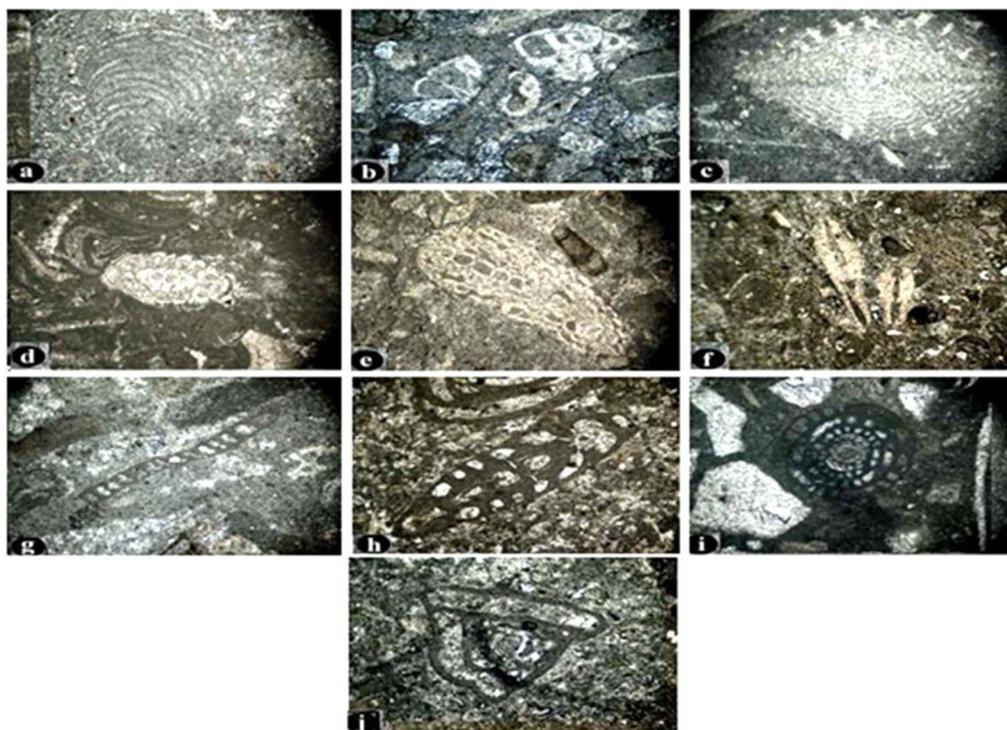


Plate-2. a. *Peneroplis evalutus* (4X) sample no. 9; b *Neorotalia viennotti* (Greig, 1935), (10X) sample no. 138; c. *Nephrolepidina* sp. (4 X) sample no. 8; d *Elphidium* sp 14 (10 X) sample no. 34; e. *Miogypsina* sp. (4 X) sample no. 9; f. *Operculina complanate* (Defrance in de Blainville, 1822) (4X) sample no. 65; g. *Meandropsina iranica* Henson, 1950 (4X) sample no. 10; h. *Dendretina rangi* d'Orbigny emend. Fornasini, 1904 (4 X) sample no. 39; i. *Borelis melo curdica* (Reichel) (4X) sample no. 7; j. *Triloculina trigonula* Lamarck, 1804 (10X) sample no: 30

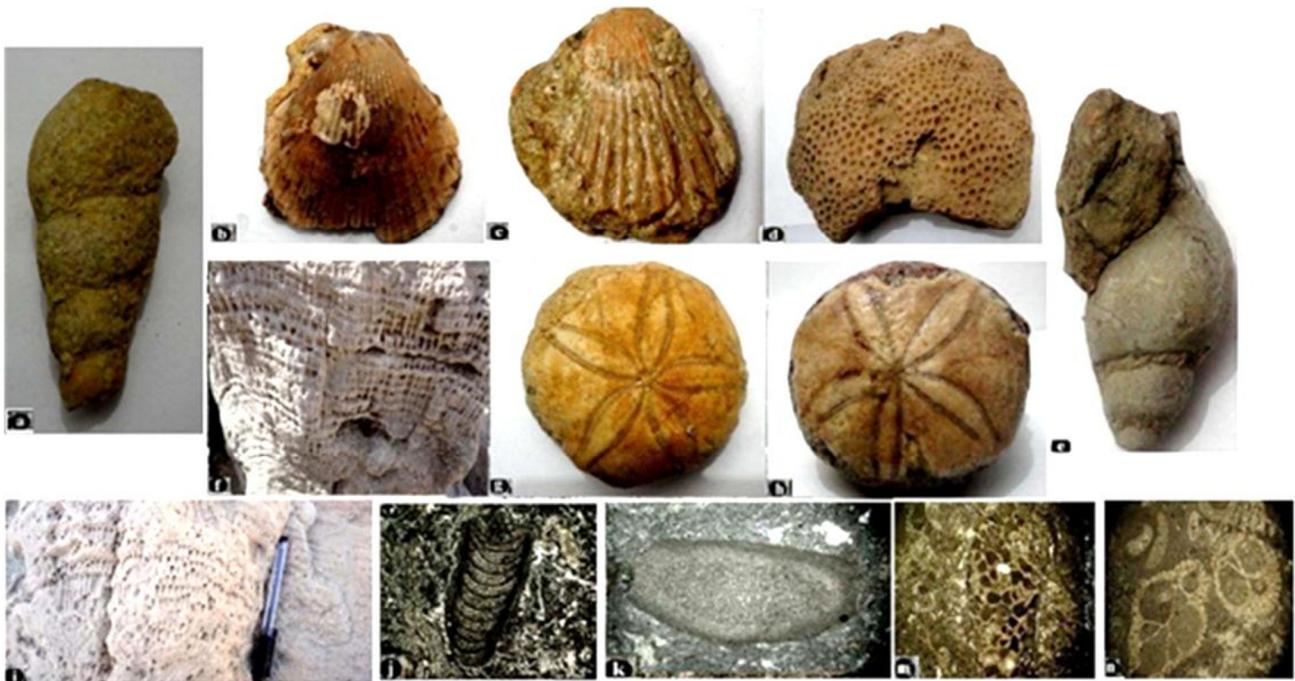


Plate-3. a. (Gastropods) *Ficus papayratium* sample No. 123 b. (Pelecypods). *Leptoplecten*cf. *L.ecnomius* Woodring, 1982; c. (Pelecypods) *Pectinidae*, sp.; d. Coral, sample no137 e. Gastropods, sample 121. f. Coral, sample No. 146; g. (Echinoemata) *Trachypatagus tuberculatus* Langhian-Serravallian, sample 143; h. (Echinoemata). *Echinolampas* cfr. *vilanovae* COTTEAU. Burdigalian, *Echinolampas (Macrolampas) discus* Desor. Aquitanian, sample No. 143; i. Coral, sample no146 j. *Lithothamnium* sp. (4X) sample no: 110, (4X); k. *Lithophyllum* sp. (4X) sample no: 75; l. *Onychocella* sp. (4X) sample no: 18. m. *Tubucellaria* sp. (10 X) sample no: 108

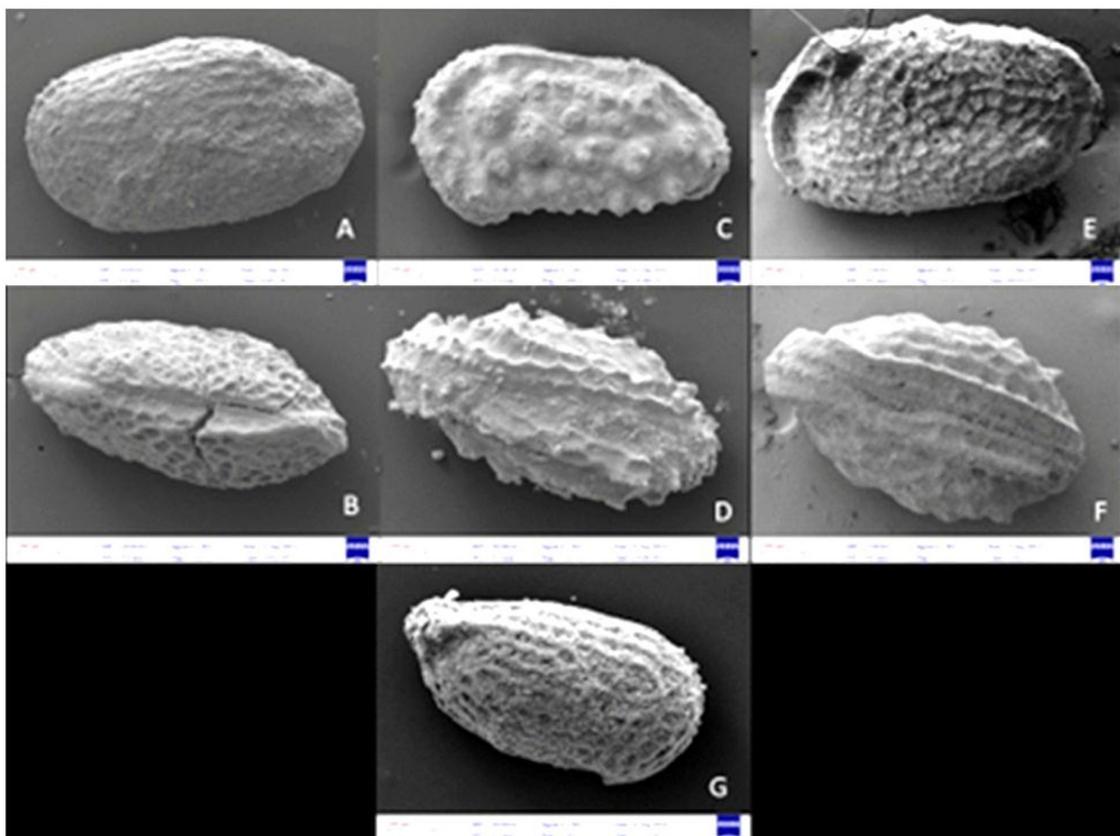


Plate 4. SEM microphotographs of selected ostracods from the Bandar-Abbas outcrop. A – *Cytheretta* sp external view; B – *Cytheretta* sp lateral view; C – *Actinocythereis rosefieldensis* (Howe & Law, 1936) left lateral view; D – *Actinocythereis rosefieldensis* (Howe & Law, 1936) lateral view; E – *Hemicythere* sp external view; F – *Hemicythere* sp lateral view; G – *Cytherura* sp external view

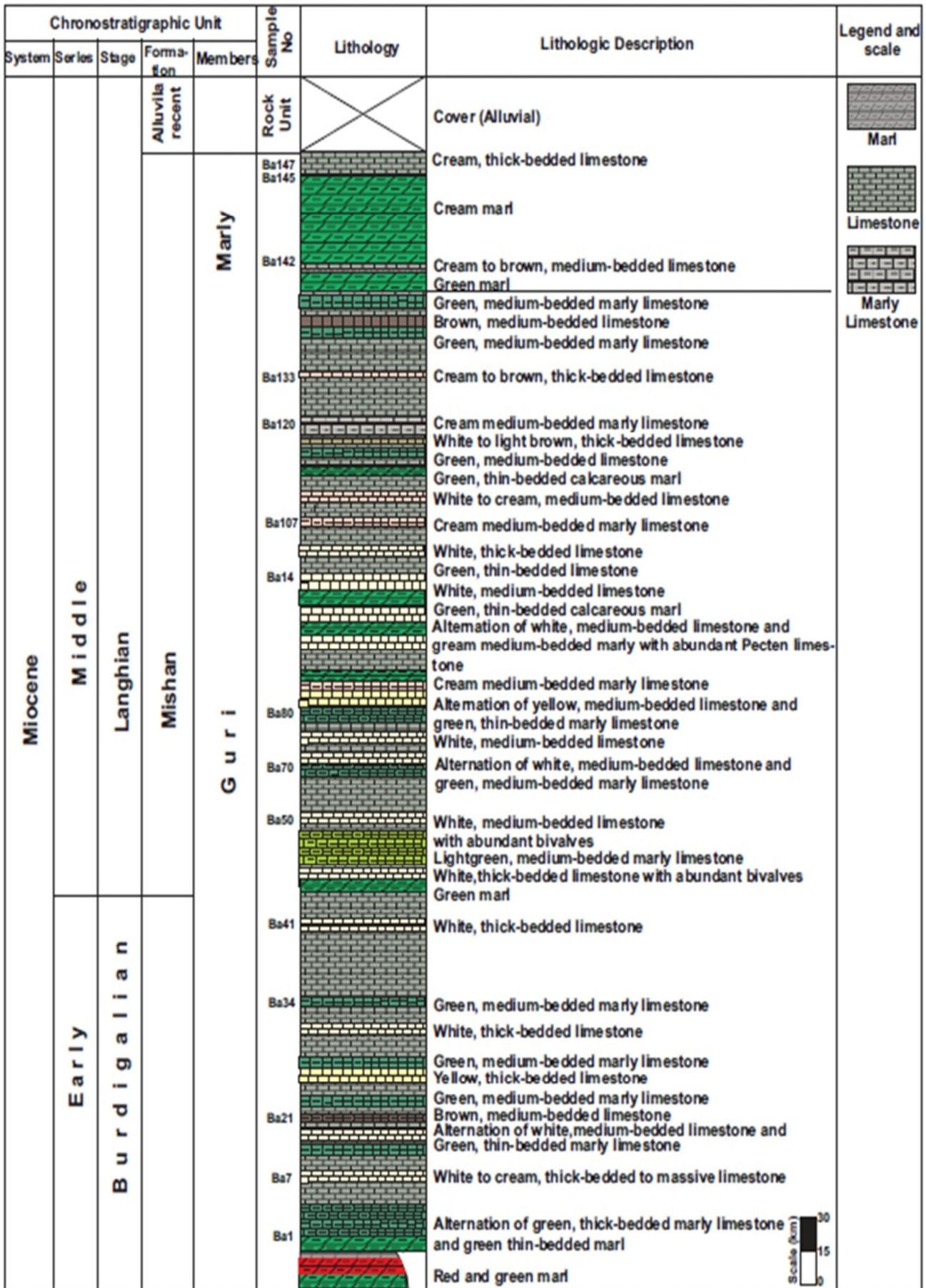


Fig. 5. Lithostratigraphic column of Early-Middle Miocene rock units in the Baghestan section of the studied area



4.1.1 *Elphidium* sp.14, *Borelis melo curdica* Interval Zone

Definition: Biostratigraphic interval between the FAD of *Elphidium* sp.14, and FAD of *Borelis melo curdica*.

Boundary zone: This zone is represented by the First Appearance Datum (FAD), and dominance of the large benthic foraminifera is represented by *Elphidium* sp. 14. Thickness: 20 meters, is recorded from the Mishan Formation, samples 1–7.

Association: It is characterized by the appearance and dominance of *Miogypsina* sp., *Schlumbergerina* sp., *Quinqueloculina* sp., *Discorbis* sp., *Neorotalia viennoti*, *Ammonia beccarii*, *Elphidium* sp.14, *Ammonia stachi*, *Triloculina trigonula*, *Amphistegina lesson*, *Amphistegina lesson*, *Nephrolepidina* sp., *Borelis melo curdica*, *Borelis haueri*, *Subterraniphyllum thomasi*, *Tubucellaria* sp., and coral.

Correlation: This zone is equivalent to the Indeterminate Zone of Laursen et al., (2009) and *Miogypsinoides* – *Archaia*s-Valvulinid Zone (*Archaia*s *asmarius*-*Archaia*s *hensoni* subzone and *Elphidium* sp.1-*Miogypsina* sp. subzone) of (Adams & Bourgeois, 1967), and to the SBZ 24 (*Austrorillina howchini*-*Miogypsina*- *miogypsinoides deharti*) of Cahuzac and Pognant, (1997), with *Austrorillina howchini*-*Peneroplis farsensis* assemblage Zone of Ahmmad (2020)) (Fig. 7). It is corresponding to the *Ammonia beccari*-*Austrorillina hawchini* zone (Ghafor and Ahmad, 2021).

Age: Early Miocene (Aquitanian).

Neogene				System
Miocene				Series
Aquitanian	Burdigalian	Langhian		Stage
<i>Archaia</i> s <i>asmarius</i> <i>Archaia</i> s <i>hensoni</i>	<i>Borelis melo</i> group- <i>Meandropsina</i> <i>iranica</i>			Adams & Bourgeois 1967
<i>Elphidium</i> sp. 14 <i>Miogypsina</i>				
<i>Miogypsinoides</i> <i>Archaia</i> s-Valvulinid				
SBZ21	SBZ5	SBZ6		Cahuzac & Pognant 1997
<i>Austrorillina</i> <i>horuchini</i> - <i>Miogypsina</i> <i>Miogypsinoides deharti</i>	<i>Borelis melo</i> group- <i>Miogypsina</i>	<i>Borelis melo</i>		
<i>Austrorillina</i> <i>horuchini</i> (AH)	<i>Borelis melo melo</i> (BMM)		<i>Borelis melo curdica</i> (BMC)	Serra-Kiel et al., 1998
<i>Miogypsina</i> <i>Elphidium</i> sp.14 <i>Peneroplis</i> <i>farsensis</i> Indeterminate	<i>Borelis melo curdica</i> <i>Borelis melo melo</i>	Not Studied		Laursen et al., 2009
<i>Elphidium</i> sp.14 <i>Miogypsina</i> sp. Zone	<i>Borelis melo curdica</i> <i>Borelis melo melo</i>			Moghadam et al., 2019
<i>Austrorillina</i> <i>horuchini</i> <i>Peneroplis</i> <i>farsensis</i>	<i>Ammonia beccari</i>	Not Studied		Ahmad 2020
<i>Elphidium</i> sp.14- <i>Borelis melo curdica</i> Interval Zone	Operculina, Amphistegina Lessoni Assemblage Zone			This Study
	<i>Borelis melo curdica</i> Total Range Zone			
	<i>Borelis melo curdica</i> - <i>Meandropsina</i> <i>Iranica</i> Assemblage Zone			
<i>Elphidium</i> sp.14- <i>Miogypsina</i> Assemblage Zone				

Fig. 7. Correlation chart showing the biostratigraphic zones of this study with the other studies



4.1.1.2 *Elphidium* sp.14, *Miogypsina* Assemblage Zone

Definition: The biostratigraphic interval of this zone is characterized by the association of taxa (*Elphidium* sp. 14 and *Miogypsina*).

Boundary zone: This zone is represented by the First Appearance Datum (FAD), and dominance of the large benthic foraminifera is represented by (*Elphidium* sp. 1 and *Miogypsina*). The Mishan Formation, samples 1–36, has a thickness of 133 meters.

Association: It is characterized by the appearance and dominance of *Schlumbergerina* sp., *Quinqueloculina* sp., *Discorbis* sp., *Neorotalia viennoti*, *Ammonia beccarii*, *Ammonia stachi*, *Triloculina trigonula*, *Amphistegina lesson*, *Nephrolepidina* sp., *Borelis melocurdica*, *Borelis haueri*, *Penetroplis* cf. *evolutus* *Meandropsina iranica*, *Dendritina rangi*, *Archaias kirkukensis*, *Amphistegina* cf. *radiat* *Subterraniophyllum thomasi*, *Lithothamnium* sp., *Tubucellaria* sp., *Memberanipora* sp., *Onychocella* sp., *Memberanipora* sp., and coral

Correlation: This zone is equivalent to the Adams & Bourgeois (1967) *Miogypsinoidea-Archaias-Valvulinid* Zone-*Borelis melo* group-*Meandropsina iranica* Assemblage Zone, Cahuzac and Poignant (1997) SBZ24 and lower part of SBZ25, Laursen et al., (2009) *Miogypsina-Elphidium* sp. It corresponds to the *Ammonia beccarii-Austritillina hawchini* zone of (Ghafor and Ahmad, 2021).

Age: Early Miocene (Aquitani–Burdigalian).

4.1.1.3 *Borelis melo curdica* – *Meandropsina iranica* Assembly Zone

Definition: The biostratigraphic interval of this zone is characterized by the association of taxa (*Borelis melo curdica* and *Meandropsina iranica*).

Boundary zone: This zone is represented by the First Appearance Datum (FAD) of *Borelis melo curdica*, and the dominance of the large benthic foraminifera is represented by *Meandropsina iranica*. A thickness of 207 meters is recorded in the Mishan Formation; samples number 7–64.

Association: It is characterized by the appearance and dominance of *Schlumbergerina* sp., *Quinqueloculina* sp., *Discorbis* sp., *Neorotalia viennoti*, *Ammonia beccarii*, *Ammonia stachi*, *Triloculina trigonula*, *Amphistegina lesson*, *Nephrolepidina* sp., *Borelis melocurdica*, *Borelis haueri*, *Penetroplis* cf. *evolutus* *Meandropsina iranica*, *Dendritina rangi*, *Archaias kirkukensis*, *Amphistegina* cf. *radiat* *Subterraniophyllum thomasi*, *Lithothamnium* sp., *Tubucellaria* sp., *Memberanipora* sp., *Onychocella* sp., *Memberanipora* sp., and coral

Correlation: This zone is equivalent to the *Borelis melo* group-*Meandropsina iranica* Assemblage Zone of Adams and Bourgeois (1967), SBZ25 and the lower part of SBZ26 of Cahuzac and Poignant (1997), the *Borelis melo* group-*Borelis melo* assemblage Zone of Laursen et al. (2009), and the *Borelis melo* group-*Borelis melo* assemblage Zone of Moghadam et al., (2019). *Ammonia beccarii* zone of Ahmmad, (2020) (Fig. 7). It corresponds to the *Ammonia beccarii-Austritillina hawchini* zone of (Ghafor and Ahmad, 2021).

Age: Early Miocene (Burdigalian–Langhian).

4.1.1.4 *Borelis melo curdica* Total Range Zone

Definition: Biostratigraphic interval between the First and Last Appearances of *Borelis melo curdica* Zone boundary: This zone is represented by the First Appearance Datum (FAD) of *Borelis melocurdica* and the last occurrence of this species. Thickness: 316 meters, is recorded in the Mishan Formation, samples 7–105.

Association: It is characterized by the appearance and dominance of *Schlumbergerina* sp., *Quinqueloculina* sp., *Discorbis* sp., *Neorotalia viennoti*, *Ammonia beccarii*, *Ammonia stachi*, *Triloculina trigonula*, *Amphistegina lesson*, *Nephrolepidina* sp., *Borelis melocurdica*, *Borelis haueri*, *Penetroplis* cf. *evolutus* *Meandropsina iranica*, *Dendritina rangi*, *Archaias kirkukensis*, *Amphistegina* cf. *radiat* *Subterraniophyllum thomasi*, *Lithothamnium* sp., *Lithophyllum* sp., *Tubucellaria* sp., *Memberanipora* sp., *Onychocella* sp., *Memberanipora* sp., and coral

Correlation: This zone is equivalent to the *Borelis melo* group *Meandropsina iranica* of Adams and Bourgeois (1967) and to the *Borelis melo curdica* total range zone of James and Wynd (1965). Van Buchem et al. (2010), and to the SBZ 26(*Borelis melo*) of Cahuzac and Poignant, (1997), with *Ammonia beccarii* Total Range Zone of Ahmmad, (2020) (Fig. 7). It corresponds to the *Dendritina rangi-Rotlia vieonti* zone (part) of (Ghafor and Ahmad, 2021).

Age: Early-Middle Miocene (Burdigalian–Langhian).

4.1.1.5 *Operculina-Amphistegina lessoni* Assemblage Zone

Definition: The biostratigraphic interval of this zone is characterized by the association of taxa (*Operculina* and *Meandropsina iranica*).

Boundary zone: This zone is represented by the First Appearance Datum (FAD) of *Operculina* and the association of the *Amphistegina lessoni*. Thickness: 337 meters, recorded from the Mishan Formation; samples number (40–147)

Association: It is characterized by the appearance and dominance of *Schlumbergerina* sp., *Quinqueloculina* sp., *Discorbis* sp., *Neorotalia viennoti*, *Ammonia beccarii*, *Ammonia stachi*, *Triloculina trigonula*, *Amphistegina lesson*, *Nephrolepidina* sp., *Borelis melocurdica*, *Borelis haueri*, *Pen-*



eroplis cf. evolutus Meandropsina iranica, Dendritina rangi, Archaias kirkukensis, Amphistegina cf. radiat Subterraniophyllum thomasi, Lithothamnium sp., Tubucellaria sp., Memberanipora sp., Onychocella sp., Memberanipora sp., and coral

Correlation: This zone is equivalent to the *Borelis melo* zone (SBZ26) of Cahuzac and Poignant, (1997) and the upper part of the *Borelis melo melo* (BMM) zone—the *Borelis melo curdica* zone of Serra kiel et al., (1998). (Fig. 7). It corresponds to the *Dendritina ranji-Rotlia vieonnti* zone (part) of (Ghafor and Ahmad, 2021).

Age: Early to Middle Miocene (Burdigalian–Langhian).

5 PALAEOECOLOGICAL INTERPRETATION

Foraminifera and non-foraminifera were used in the paleoecology of Paleogene-Neogene carbonate platforms (Hottinger, 1997; Pedley, 1998; Geel, 2000; Pomar, 2001; Romero et al., 2002; Cosovic et al., 2004; Pomar and Hallock, 2008; Ghafor and Ahmad, 2021; Ghafor et al., 2023; Ghafor et al., 2023). Bou-Dagher-Fadel, 2008), clarified that the shapes and sizes of the foraminiferal species are significant and occur in many different environments, from near-shore to the deep-sea environment. The recorded biozones were calibrated with the previously studied Miocene planktonic foraminiferal biozones on the same samples and correlated with the Miocene redeposited ostracods regardless of their limited presence in the studied outcrop. These ostracods are also found in the neighbouring countries in Miocene deposits. The calcareous red algae are also used in palaeoecological studies. Many parameters and variables influence foraminiferal distribution in aquatic life: temperature, water depth, hydrostatic pressure, light intensity, sediment type, current systems, salinity, nutrients, and oxygen (Table 1).

Table 1 – Vertical distribution of faunal composition, palaeoecological elements (Temperature, light, depth, hydrodynamic energy, substrate, salinity, nutrient, and oxygen estimation) of the Guri Member (Mishan Formation) in the studied section

Age	Early-Middle Miocene
Fossil Content Families	<i>Sortidae, Peneroplidae, Miliolidae, Rotalidae, Amphestedae, Hauerinidae, Alveolinidae, Meandropsinidae, Elephinidae, Miogypsinidae, Miogypsinidae.</i> Non-Foraminifera, algae bryozoans' molluscan macrofossils -bivalves and gastropods.
Temperature	Subtropical to tropical
Light	Mesophotic to euphotic
Depth	Open marine environments to shallower water depths
Hydrodynamic energy	Large shells with thin walls in open marine foraminifera and smaller and thicker shell in shallow and restricted environments.
Substrate	Soft sediments to hard sediments
Salinity	Normal to higher salinity environments ranges from 34 to 50 ‰.
Nutrients and oxygen	Mesotrophic to oligotrophic

5.1 Temperature: Temperature acts as a barrier to the dispersal of marine animals. (Brasier, 2004; Wilson and Vecsei, 2005) shows that larger benthic foraminifera live in tropical and subtropical water environments and restrict to temperatures of about 14 to 25 °C. According to Hattinger, (1997), most of the perforate foraminifera such as *Nummulites, Miogypsina, Operculina,* and *Amphistegina* are abundant in subtropical to tropical environments, while some imperforate foraminifera such as *Archaias, Borelis, Peneroplis,* and miliolids are restricted to tropical environments (BouDagher-Fadel and Wilson, 2000; Brandano et al., 2009); and (Fügel, 2010; Brandano et al., 2010) show that red algae such as *Lithoporella, Lithium,* and *Lithotamini-um* are restricted in subtropical to tropical environments. Therefore, the presence of perforate large benthic foraminifera such as *Operaculina* and *Amphistegina* in the middle to the upper part of the Mishan Formation in the studied section reflects subtropical to tropical environments, and the presence of imperforate large benthic foraminifera such as *Archaias, Dendritina, Peneroplis,* and miliolids in the lower to the middle part of the Mishan Formation at the studied section shows a tropical environment.

5.2 Light: While light penetration in the oceans (the photic zone) is attractive to foraminifera (Brasier 2004), the larger benthic foraminifera such as *Amphistegina, Heterostegina,* and *Operculina* reflect mesophotic to oligophotic conditions (Pomar, 2001; Hohenegger 2004; Beavington-Penney and Racey 2004; BouDagher-Fadel, 2008). The change in the test shape reflects decreased light levels at greater depths (BouDagher-Fadel, 2008). It is known that larger foraminifera living in fairly turbulent waters become relatively large, with a thickness-to-diameter ratio of 0.6–0.7 (Cosovic et al., 2004). Associations of larger foraminifera and symbiotic red algae are well-dependent on the photic zone (Barattolo et al., 2007). The abundance of larger benthic foraminifera such as *Borelis, Peneroplis, Dendritina, Archaias,* and zooxanthellate corals in the studied section of the Mishan Formation indicate a mesophotic to euphotic condition (Halfar et al., 2004; Pomar and Hallock, 2008; Brandano et al., 2009b; Mazzucchi and Tomassetti, 2011; Tomassetti and Brandano, 2013). Larger perforate foraminifera is represented by *Operculina, Elphidium,* and *Miogypsina* in



the Guri Member of the Mishan Formation, and comparison with analogs in the modern platform led to the interpretation of these sediments as being photic zone deposits (Hottinger, 1980; Hottinger, 1983; Leutenegger, 1984; Reiss, and Hottinger, 1984; Hohenegger, 1996; Hallock, 1999).

5.3 Depth: According to Brassier (2004), foraminifera is found from sea level to more than 10,000 meters. Changes in water depth could affect environmental changes such as sedimentation rate, turbulence, light intensity, organic change, hydrodynamic energy, and dissolved oxygen (Leckie and Olsen, 2003). Large Benthic Foraminifera (LBF) was previously used to determine water depth due to its sensitivity to it. Some larger benthic foraminifera, such as *Amphistegina* and *Nummulites*, become flatter with thinner outer walls with increasing water depth and decreasing light intensity. This reveals the dependence of morphology on the depth gradient (Pecheux 1995). Some imperforate benthic foraminifera such as *Archaias*, *Peneroplis*, and *Borelis* are indicators of shallow waters below 30 m (Romero et al., 2002; Murray, 2006). Most of the perforate foraminifera, such as *Nummulites*, *Miogypsina*, *Operculina*, and *Amphistegina*, are distributed in a water depth of about 40–70 m (Hottinger, 1997); according to Flügel (2010; Brandano et al. (2010), red algae, such as *Lithoporella*, *Lithophyllum*, and *Lithotaminium*, are restricted to a depth of 20–80 m. (Bou-Dagher-Fadel and Wilson, 2000; Brandano et al., 2009); and (Flügel, 2010; Brandano et al. 2010). *Operculina*, *Elphidium*, and *Miogypsina* in the Guri Member of the Mishan Formation are the most important indicators for warm, shallow marine environments (Geel, 2000; Sajadimet et al., 2014). At the lower part of the Mishan Formation, perforate foraminifera are common and reflect an open marine environment, while in the lower and middle part of the studied section, imperforate foraminifera such as *Peneroplis* and miliolids are abundant and generally found in shallower water (20–80 m)

5.4 Hydrodynamic energy: Depending on hydrodynamic water energy, some foraminifera develop thick lamellar crust shells, while others tend to develop a hard, conical shell shape with abundant secondary cells (Rasser et al., 2005). *Operculina* and *Heterostegina* have inhabited environments ranging from high-energy, shallow-water fore reef facies (BouDagher-Fadel 2008) to quiet waters near the base of the photic zone (Chaproniere, 1975). In the studied area, open marine foraminifera such as *Operculina*, *Nerhrolepidina*, *Discocyclina*, and *Amphistegina* have large shells with thin walls, while those with smaller and thicker shell walls live in shallow and restricted environments.

5.5 Substrate: Foraminifera from coarser substrates tend to be thick-shelled, heavily ornamented forms of lenticular or globular shape. Low-energy habitat with fine-grained substrates is attractive to many faunal species with thin, delicate, and elongated shells (Brasier 2004). Conical and thick-shelled forms of foraminifera prefer to live on coarse-grained substrates, while flattened forms prefer soft substrates (Brasier, 1975). *Operculina* is inhabited on soft sediment, while *Borelis* and *Archaias* live in relatively unstable substrates in shallow environments (Geel, 2000). *Archaias* and *Operculina* can live in environments with about 40 % clastic influx but increases in clastic influx led to a reduction in their numbers (Kumar and Saraswati, 1997). So, the Mishan Formation of the studied section has deposited soft to hard sediments.

5.6 Salinity: A miliolid-dominant environment in a benthic foraminifer assemblage reflects decreased circulation and reduced oxygen content, or euryhaline conditions. Miliolids are found in a variety of very shallow, hyposaline to hypersaline environments and are also common in sand shoal environments of normal salinity (Brasier, 1975a). They are generally evidence of a restricted lagoon environment (Wilson, 1975). Perforate-wall foraminifera such as *Nummulites*, *Operculina*, and *Amphistegina* wall refer to normal seawater conditions with salinity ranges from 34 to 40 (Geel 2000; Mossadegh et al., 2009). The association of imperforate foraminifera such as *Archaias* and *Peneroplis* with coral and red algae reflects a salinity of 40 to 50 (Mossadegh et al., 2009). As a result, the middle to the upper part of the Mishan Formation in the studied section has deposited in a normal water salinity environment (with salinity ranges from 34 to 40), while the lower to the middle part of the studied section experienced intermittent normal and higher salinity environments (with salinity ranges from 40 to 50).

5.7 Nutrients and oxygen: If the food supply is low, as in the deep sea, foraminiferal densities tend to be low, but diversity can be high. However, if the food supply is high, foraminiferal diversity tends to be low. (Brasier, 2004). The imperforate foraminifera, such as *Archaias* and miliolids, indicate low levels of nutrients (Geel, 2000; Samankassou, 2002; Romero et al., 2002). Also, coral reefs develop in submarine environments with the lowest levels of nutrients (Schlager 2005). Increasing amounts of nutrients lead to thriving red algae, and decreasing amounts of nutrients cause coral growth and development (James et al., 1999). Nutrient conditions have a negative relationship with temperature (Samankassou, 2002). The presence of larger benthic foraminifera indicates mesotrophic to oligotrophic conditions (Geel, 2000; Romero et al., 2002). The occurrence and abundance of large benthos foraminifera such as *Amphistegina*, *Peneroplis*, *Operculina*, and *Archaias*, along with coral and red algae in the

6 DISCUSSIONS

All the previous biostratigraphy and palaeoecological studies of the Mishan Formation did not explain it in detail (Wynd, 1965; Adams et al., 1967; Rashidi et al., 2015; Amirshahkarami and Taheri, 2009; Joudaki and Baghbani, 2018), but the present interpretation in the studied section shows that more than 30 species of foraminifera and non-foraminifera have been identified and subdivided into five biozones that the Early - Middle Miocene age was recorded of the Guri Member (Mishan Formation), and the interpretation of the pal-



aeoecological results, gives us the following results, the Member was deposited from subtropical to tropical environments, mesotrophic to oligotrophic conditions, soft sediments to hard sediments, open marine to shallower water depths, and of a normal water salinity environment ranging from 34 to 50 ‰. Ostracods presented in the lower and upper parts of the studied section indicated an inner neritic marine environment of moderate energy of currents and rapid sedimentation, while the assemblages in the middle part showed deeper (outer neritic) environments with low energy of currents and low rate of sedimentation. In general, ostracodal biostratigraphy is not as detailed as those based on foraminifera, due to their great sensitivity to ecological factors that control their distribution.

7 CONCLUSIONS

The foraminiferal contents for the same rock samples with ostracods and other macrofossils were employed in age determination and correlation. High-resolution biostratigraphy and palaeoecological investigations were carried out on the Guri Member in the Bandar Abbas area of the Zagros Mountains using mainly benthic foraminifera. In this study, 31 diagnostic benthic foraminifera species of the 17 genera, were encountered, and seven non-foraminifera with macrofossils of Mollusca, Echinodermata, bryozoans, seven ostracods, and corals were distributed, which led to five zones, revealing the early-middle Miocene age. The palaeoecological study shows that the Guri Member of the Mishan Formation in the studied section was deposited under subtropical to tropical environments, from mesotrophic to oligotrophic conditions, with soft sediments to hard sediments from open marine environments to shallower water depths, and deposited in normal water salinity environment ranging from 34 to 50 ‰.

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